

NOVEL ANTINEMATODAL AND ANTIPARASITIC AGENTS FROM
PENICILLIUM CHARLESII

II. STRUCTURE DETERMINATION OF PARAHERQUAMIDES B, C, D, E, F, AND G

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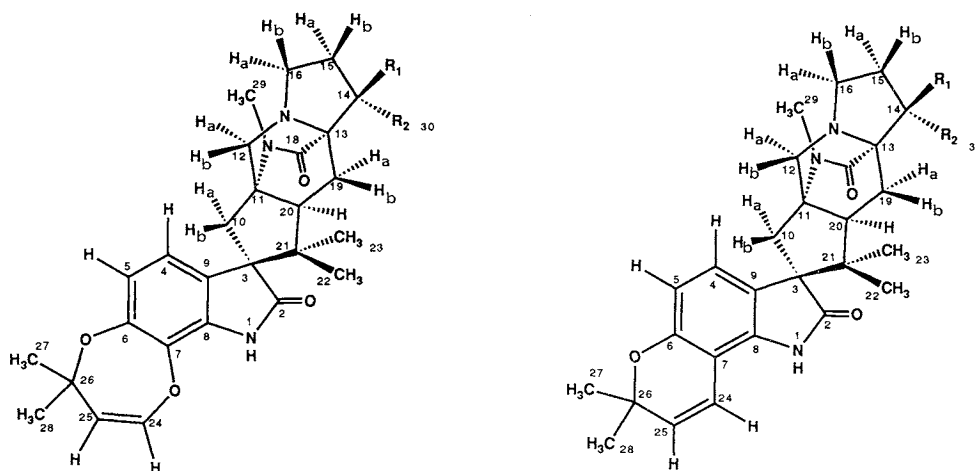
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Paraherquamides B (2, $C_{27}H_{33}N_3O_4$), C (3, $C_{28}H_{33}N_3O_4$), D (4, $C_{28}H_{33}N_3O_5$), E (5, $C_{28}H_{35}N_3O_4$), F (6, $C_{28}H_{35}N_3O_3$), and G (7, $C_{28}H_{35}N_3O_4$) are novel metabolites of *Penicillium charlesii*. The structures of these compounds have been determined by NMR and MS analysis.

The fermentation, isolation, and characterization of six novel analogs (2~7) of paraherquamide¹ (1) have been reported in the preceding paper². In this paper, the structure determination of these compounds is described.

Results and Discussion

The structures of the present compounds were determined based upon interpretation of their NMR and MS and comparison with previously reported paraherquamide¹ (1). Paraherquamide (1) has the MW 493 ($C_{28}H_{35}N_3O_5$) and exhibits critical fragment ions at m/z 434, 165, and 163 in its EI-MS. The fragment ion at m/z 434 corresponds to neutral loss of *N*-methylformamide from the molecular ion. Cleavage of bonds C-20~C-21 and C-10~C-3 (see 1) with loss of *N*-methylformamide affords m/z 163, the saturated



- 1 $R_1=OH$ $R_2=CH_3$
 2 $R_1=H$ $R_2=H$
 3 $R_1, R_2=H_2C=$
 4 $R_1, R_2=-OCH_2-$
 5 $R_1=H$ $R_2=CH_3$

- 6 $R_1=H$ $R_2=CH_3$
 7 $R_1=OH$ $R_2=CH_3$

Table 1. ^1H NMR data for paraherquamides (1~7)^a.

Proton	1 ^b	1	2	3
4-H	7.03 d (8.1)	6.84 d (7.3)	6.85 d (8.2)	6.86 d (8.2)
5-H	6.66 d (8.1)	6.68 d (7.3)	6.67 d (8.2)	6.68 d (8.0)
10-Ha	2.67 d (15.3)	2.65 d (16.1)	2.67 d (15.2)	2.68 d (15.2)
10-Hb	1.97 d (15.4)	1.87 d (15.6)	1.87 d (15.1)	1.88 d (15.4)
12-Ha	2.58 dd (1.8, 11.4)	2.55 dd (2.2, 11.0)	2.61 dd (1.0, 11.5)	2.68 dd (1.2, 11.2)
12-Hb	3.65 d (11.4)	3.58 d (10.8)	3.59 d (10.9)	3.63 d (10.7)
15-Ha	1.75 ^c dd	1.80 ^c m	1.39 ^c m	2.55 ^c m
15-Hb	2.18 ^c m	2.24 ^c m	2.13 ^c m	2.55 ^c m
16-Ha	2.14 ^c m	2.18 ^c m	2.13 m	2.29 ^c m
16-Hb	3.15 ^c ddd	3.17 ^c ddd	3.03 m	3.06 ^c m
19-Ha	1.83 dd (10.3, 12.9)	1.77 dd (10.8, 10.8)	1.83 m	2.03 dd (11.4, 12.3)
19-Hb	1.71 dd (11.0, 12.9)	1.75 dd (10.5, 12.5)	1.64 dd (10.4, 12.4)	1.65 dd (9.5, 12.3)
20-H	3.00 ddd (1.7, 10.5, 10.5)	2.96 ddd (2.0, 10.3, 10.3)	2.98 m	3.06 m
22-H	1.11 s	1.08 s	1.09 s	1.09 s
23-H	0.82 s	0.84 s	0.83 s	0.86 s
24-H	6.36 d (7.7)	6.32 d (7.8)	6.32 d (7.7)	6.33 d (7.7)
25-H	4.97 d (7.8)	4.90 d (6.8)	4.89 d (7.7)	4.90 d (7.7)
27-H	1.39 ^d s	1.41 ^d s	1.41 ^d s	1.41 ^d s
28-H	1.42 ^d s	1.43 ^d s	1.43 ^d s	1.44 ^d s
29-H	2.99 s	2.99 s	3.01 s	3.01 s
14-OH	2.75 br s	2.66 br s	—	—
1-H	9.45 br s	7.50 br s	7.41 br s	7.41 br s
14-H	—	—	1.83 ^d m, 2.49 ^d m	—
30-H	1.54 s	1.56 s	—	4.97 m (<2), 5.13 m (<2)

Proton	4	5	6	7
4-H	6.86 d (8.2)	6.84 d (8.5)	6.94 d (8.4)	6.93 d (8.5)
5-H	6.68 d (8.2)	6.67 d (7.5)	6.42 d (8.3)	6.42 d (8.2)
10-Ha	2.66 d (15.4)	2.66 d (15.9)	2.64 d (15.0)	2.64 d (15.6)
10-Hb	1.89 d (15.4)	1.86 d (16.0)	1.83 d (15.8)	1.85 d (15.7)
12-Ha	2.61 dd (1.4, 11.2)	2.51 br d (10.6)	2.50 dd (1.0, 10.7)	2.56 dd (1.5, 11.5)
12-Hb	3.67 d (10.9)	3.56 br d (10.7)	3.54 d (11.4)	3.58 br d (11.2)
15-Ha	2.36 ddd (1.5, 8.8, 13.6)	1.75 dddd (4.5, 10.7, 10.7, 10.7)	1.75 dddd (4.5, 10.6, 10.6, 10.6)	1.79 m
15-Hb	1.92 ddd (8.9, 8.9, 13.7)	1.97 m	1.97 m	2.25 m
16-Ha	2.27 ddd (8.6, 8.6, 8.6)	2.20 m	2.20 ddd (5.5, 8.9, 10.6)	2.20 m
16-Hb	3.22 ddd (1.5, 9.0, 9.0)	3.13 m	3.12 ddd (4.4, 9.0, 9.0)	3.18 ddd (4.5, 9.0, 9.0)
19-Ha	1.57 dd (9.9, 12.4)	2.00 dd (11.0, 11.2)	2.00 dd (11.1, 11.5)	1.82 dd (10.6, 12.8)
19-Hb	1.41 m	1.38 m	1.38 m	1.76 dd (10.4, 12.9)
20-H	2.96 m	2.96 m	2.98 m	2.98 ddd (1.5, 10.8, 10.8)
22-H	1.10 s	1.08 s	1.07 s	1.09 s
23-H	0.82 s	0.84 s	0.83 s	0.84 s
24-H	6.32 d (7.7)	6.33 d (8.2)	6.34 d (9.6)	6.40 d (9.8)
25-H	4.90 d (7.7)	4.90 d (8.0)	5.77 d (9.7)	5.73 d (9.9)
27-H	1.41 ^d s	1.41 ^d s	1.41 ^d s	1.41 ^d s
28-H	1.43 ^d s	1.43 ^d s	1.44 ^d s	1.44 ^d s
29-H	3.01 s	2.97 s	2.97 s	3.00 s
14-OH	—	—	—	2.69 br s
1-H	7.41 br s	7.57 br s	8.32 br s	8.96 br s
14-H	—	1.89 m	1.89 m	—
30-H	2.93 d (4.3), 3.08 d (4.3)	1.36 d (6.8)	1.37 d (6.9)	1.57 s

^a Spectra recorded at 400 MHz in CD_2Cl_2 except where noted. Chemical shifts in ppm referenced to CD_2Cl_2 at 5.32 ppm as internal standard. Data in parentheses are coupling constants in $J=\text{Hz}$. ddd = doublet of doublet of doublets. dddd = doublet of doublet of doublet of doublets.

^b Spectrum taken in acetone- d_6 , referenced to solvent peak at 2.04 ppm as internal standard.

^c Spin system showing second order effects, observed coupling constants are not accurate.

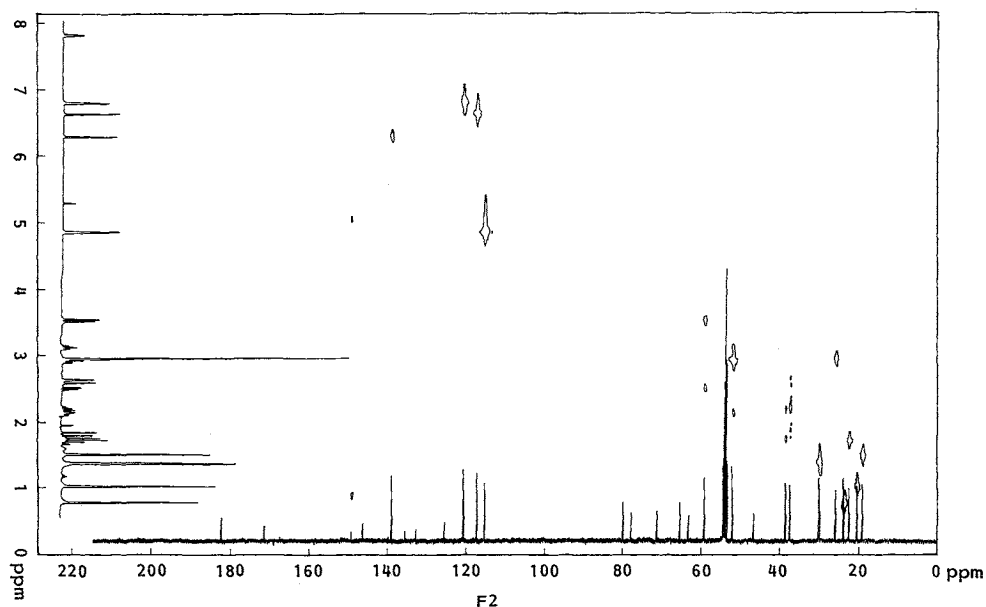
^d Interchangeable assignments.

Table 2. ^{13}C NMR data for **1**.

Carbon	δ (ppm)	m	$^1J_{\text{C-H}}$	Carbon	δ (ppm)	m	$^1J_{\text{C-H}}$
C-2	182.34	s	—	C-16	52.04	t	140
C-3	63.32	s	—	C-18	171.47	s	—
C-4	120.73	d	161	C-19	22.53	t	133
C-5	117.31	d	162	C-20	51.97	d	130
C-6	146.23	s	—	C-21	46.62	s	—
C-7	135.48	s	—	C-22	20.59	q	126
C-8	132.71	s	—	C-23	23.91	q	126
C-9	125.48	s	—	C-24	139.06	d	193
C-10	37.44	t	131	C-25	115.30	d	155
C-11	65.55	s	—	C-26	80.00	s	—
C-12	59.27	t	135	C-27	29.92	q	127
C-13	71.46	s	—	C-28	30.06	q	128
C-14	78.03	s	—	C-29	25.96	q	129
C-15	38.57	t	131	C-30	19.17	q	138

Chemical shifts in ppm referenced to CD_2Cl_2 at 53.8 ppm as internal standard. All coupling constants are in $J=\text{Hz}$. Carbon numbering is based on that of the marfortines³⁾.
m: Multiplicity.

Fig. 1. One-bond ^{13}C - ^1H chemical shift correlation (HETCOR) NMR spectrum for paraherquamide (**1**) at 19°C in CD_2Cl_2 at 100 MHz.



fused ring portion of the molecule. The lower intensity m/z 165 ion appears to result from loss of methylisocyanate ($\text{CH}_3\text{N}=\text{C}=\text{O}$) rather than *N*-methylformamide.

The published ^1H NMR spectrum¹⁾ of **1** in CDCl_3 is complicated by the poor spectral characteristics in this solvent. Subsequent ^{13}C and ^1H NMR studies with **1** in CD_2Cl_2 and $(\text{CH}_3)_2\text{CO}-d_6$ led to complete assignment of all the resonances (Tables 1 and 2). In particular, ^{13}C - ^1H correlation experiments (HETCOR, Fig. 1) were critical in definitively assigning the following proton resonances on the

resonances together with the absence of the C-14-methyl resonance observed for **1** suggested the presence of a spiro-epoxide moiety. The location of the epoxide on the pyrrolidine ring was confirmed by analysis of the remaining resonances. A four spin system consisting of resonances at 1.92 (15-Hb), 2.36 (15-Ha), 2.27 (16-Ha) and 3.22 (16-Hb) ppm was assigned to the methylene protons on carbons 15 and 16. The relative stereochemistry of the protons was assigned on consideration of the coupling constants and ring geometry. The $J=1.5$ Hz coupling between the 3.22 (16-Hb) and 2.36 (15-Ha) ppm resonances implied a near 90 degree dihedral angle between these protons. This established a $J=9$ Hz geminal coupling between the 3.22 (16-Hb) and 2.27 ppm (16-Ha) resonances and a $J=13.7$ Hz geminal coupling between the 2.36 (15-Ha) and 1.92 ppm (15-Hb) resonances. Both these values are in good agreement with observed literature geminal coupling constants for pyrrolidine ring systems⁴. ¹³C NMR data including Attached proton test⁵, used to establish carbon multiplicities confirmed the structural assignment. In comparison with paraherquamide, the carbon data indicated an upfield shift for C-15 from 38.57 to 31.09 ppm and for C-14 from 78.03 to 63.00 ppm, and replacement of the C-30 (19.17 ppm) methyl signal with a methylene resonance at 46.48 ppm.

Paraherquamide E (**5**) has the molecular formula $C_{28}H_{35}N_3O_4$ by HR-MS and this formula is O (oxygen) less than that of **1**. A strong fragment ion at m/z 418 which corresponds to $(M-59)^+$ is observed suggesting the presence of the standard *N*-methylamide moiety. The critical ions at m/z 147 and 149 reflect the 16 mass unit difference from **1** and indicate that the oxygen is missing from the saturated fused ring portion of the molecule, presumably from C-14. The ¹H NMR spectrum obtained for **5** was quite similar to that of **1**. The critical differences noted were the absence of the C-14-hydroxyl resonance, the appearance of a new methine multiplet at 1.89 ppm (14-H), and the change in the C-14-methyl resonance from a singlet at 1.56 ppm to an upfield shifted doublet ($J_{H-H}=6.8$ Hz) at 1.36 ppm. These observations allowed structural assignment of **5** as the C-14-deshydroxy derivative of **1**.

The molecular formula $C_{28}H_{35}N_3O_3$ was determined for paraherquamide F (**6**) by HR-MS and this formula is O₂ less than that of **1**. The standard *N*-methylamide moiety is indicated by the strong $(M-59)^+$ at m/z 402. The critical ions at m/z 147 and 149 indicate that one oxygen is missing from the saturated fused ring portion of the molecule, presumably from C-14, and by difference, the second oxygen equivalent must be missing from the indolone portion of the molecule. The ¹H NMR spectrum showed the absence of the C-14-hydroxyl resonance and the change in the C-14-methyl resonance (1.56 ppm singlet to 1.37 ppm doublet coupled to 1.89 ppm (14-H) methine multiplet) as observed for **5**. In addition, the spectrum of **6** exhibited a downfield shift of the C-25 olefinic proton resonance from 4.90 to 5.77 ppm with a change in the 24-H~25-H coupling constant from $J=7.8$ to 9.7 Hz. These changes are consistent with deletion of the oxygen α to the C-24~C-25 double bond to form a 2,2-dimethyl- α -pyran ring as in **6** rather than the dihydro-1,4-dioxepin of **1**.

Paraherquamide G has the molecular formula $C_{28}H_{35}N_3O_4$ by HR-MS and this formula is O (oxygen) less than that of **1**. The characteristic $(M-59)^+$ ion suggests the standard *N*-methylamide moiety. The critical ions observed at m/z 163 and 165 as in **1**, suggest by difference that the oxygen must be missing from the indolone portion of the molecule. The ¹H NMR data for **7** displayed a close similarity to **1** except for the C-25 olefinic proton which was shifted downfield to 5.73 ppm with a concomitant change in the 24-H~25-H coupling constant as observed in **6**, again indicating a 2,2-dimethyl- α -pyran ring rather than the dihydro-1,4-dioxepin of **1**.

The relative stereochemistry of the above compounds was established *via* pure-absorptive mode 2D

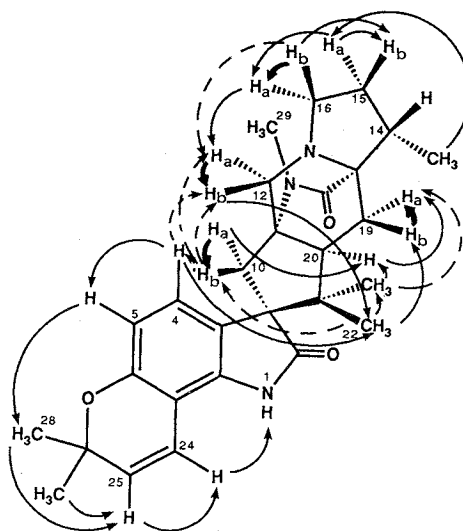
Table 3. EI-MS of compounds 2~7 (m/z (relative intensity)).

2	463.2488 (4, M^+ , $C_{27}H_{33}N_3O_4$), 404 (78), 135 (48), 133 (100)
3	475.2476 (8, M^+ , $C_{28}H_{33}N_3O_4$), 416 (30), 147 (40), 145 (100)
4	491.2436 (68, M^+ , $C_{28}H_{33}N_3O_5$), 432 (80), 163 (36), 161 (100)
5	477.2602 (2, M^+ , $C_{28}H_{35}N_3O_4$), 418 (48), 149 (36), 147 (100)
6	461.2665 (2, M^+ , $C_{28}H_{35}N_3O_3$), 402 (42), 149 (36), 147 (100)
7	477.2612 (2, M^+ , $C_{28}H_{35}N_3O_4$), 418 (24), 165 (28), 163 (100)

NOE experiments on several of the structural analogs (4~7). A representative data set is presented for 7 in Fig. 3. The medium intensity NOE's from 12-Hb and 4-H to the 22-H methyl group were critical in establishing the relative stereochemistry of the spiro center. The NOE data also allowed stereo-specific proton assignments of the geminal proton pairs for 10-H, 12-H, 15-H, 16-H, 19-H and 20-H, and one set of geminal methyl groups (22-H and 23-H). Similar sets of NOE results to those presented for 7 in Fig. 3 were obtained for the analogs, indicating the same relative stereochemistry as 7. Extrapolation to the other analogs and paraherquamide led to the proton assignments as in Table 1 and the relative stereochemistries shown for 1~7. The NOE determined stereochemistry is in agreement with the stereochemistry established by the recently published crystal structure of a synthetic paraherquamide analog⁶⁾ and the crystal structure of paraherquamide¹⁾.

Fig. 3. NOE data in CD_2Cl_2 for paraherquamide G (7).

The figure depicts 1H - 1H NOE's of weak (<2.5% of diagonal based on volume integration of diagonal and cross peak), medium (2.5~10%), and strong (>10%) intensities as dashed, solid, and bold arrows, respectively.



Experimental

1H NMR spectra were recorded at ambient room temperature in CD_2Cl_2 on a Varian XL-400 NMR spectrometer at 400 MHz. Chemical shifts are shown in ppm relative to TMS at 0 ppm using the solvent peak at 5.32 ppm as internal reference. ^{13}C NMR spectra were recorded in CD_2Cl_2 at ambient room temperature on a Varian XL-400 spectrometer at 100 MHz using Waltz-16 proton decoupling. Chemical shifts are given in ppm relative to TMS at 0 ppm using the solvent peak at 53.8 ppm as internal reference. 1H - 1H chemical shift correlation spectra for 1~7 (COSY): Spectra were recorded in CD_2Cl_2 or $(CH_3)_2CO-d_6$ using the standard pulse sequence⁷⁾. Typically, a $2K \times 2K$ data set was accumulated in 512 increments with 16 or 32 transients for each value of t_1 . The delay time between scans was 1.0 second. 1H - ^{13}C chemical shift correlation spectrum for 1 (HETCOR): Spectra were recorded in CD_2Cl_2 using the standard pulse sequence⁸⁾. A $512 \times 2K$ data set was accumulated in 128 increments with 400 transients for each value of t_1 . The delay time between scans was 1.0 second and the experiment was optimized for $^1J_{CH} = 140$ Hz. The related experiment was used to establish long-range connectivities, optimizing for a multiple bond ^{13}C - 1H coupling constant of 7 Hz. The $512 \times 2K$ data set was accumulated as above with 800 transients for each value of t_1 . Pure-absorptive mode 2D NOE effect for 4~7 (NOESY): Spectra were recorded in CD_2Cl_2 for dilute (approximately 2.5 mg in 0.5 ml), degassed samples using the standard pulse

sequence with phase-sensitive detection⁹). Typically $2K \times 2K$ data sets were accumulated in 256 increments with 192 transients for each value of t_1 . Mixing time was 0.5 second and the delay time between scans (equilibration delay) was 2.5 seconds.

MS were recorded on a Finnigan-MAT MAT212 instrument by EI at 90eV. Exact mass measurements were made on the same instrument at HR by the peak matching method using perfluorokerosene as the internal standard.

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